

Global Polio Laboratory Network



For safe handling and storage of type 2 poliovirus (PV2) in GPLN laboratories

Document version (date)	Description of substantive revisions
Version 3 (March	 Adaptation of the Guidance following the Containment Advisory Group
2018)	(CAG) decision regarding handling of PV2 RNA



Scope

This document is intended to describe changes in the handling and storage of type 2 viruses within the Global Polio laboratory Network. The rationale is that by <u>31st of July 2016</u> all polioviruses type 2 (PV2) and biological materials potentially infectious for PV2 must be destroyed or contained (onsite or after transfer to designated/certified Poliovirus essential facilities).

As diagnostics laboratories, the polio laboratories will continue to receive stool samples from AFP cases and sewage samples from Environmental Surveillance, as well as other biological materials, which may contain polioviruses mainly during the first 3 to 4 months after the last day of the worldwide coordinated OPV2 withdrawal from the trivalent OPV. Therefore specific measures must be implemented to avoid unintentional release of poliovirus in the environment by minimizing polio laboratory-associated risk.

Therefore this guidance paper focuses on:

- (i) better definition of the Polio Laboratories duties based on their capacities to perform WHOrecommended testing procedures for polio diagnosis i.e Isolation, Intratypic Differentiation and Sequencing of polioviruses, and
- (ii) practical steps to be taken when a Laboratory come across a type 2 poliovirus during the diagnostic process.

It summarizes dispositions, roles and responsibilities of Laboratories and WHO to ensure standard handling and storage practices for PV2 within the GPLN starting 1^{st} August 2016.

This third version of the Guidelines (published in December 2017) aim to adapt the guidance to reflect Containment Advisory Group (CAG) decision regarding handling and storage of PV2 RNA. Indeed, following CAG recommendations, an addendum to GAP-III (containing the chapter below) will be published. The main change brought by decision is that Polio laboratories which have sequencing capacities but are not Polio Essential Facilities, are allowed to inactivate the poliovirus isolates and perform sequencing on extracted nucleic acids.

Addendum to Annex-1 of GAP-III:

Poliovirus nucleic acid: RNA, cDNA and total nucleic acid, extracted/purified from poliovirus infectious materials (e.g., a virus isolate) or potentially infectious materials (e.g., stool, respiratory specimen, sewage) using methods demonstrated to inactivate poliovirus, or synthesized poliovirus RNA or cDNA RNA/cDNA (e.g., cDNA clone, synthetic transcript) can be handled outside of poliovirus containment under the condition that these materials will not be introduced into polio-permissive cells or animals as described in this standard and the 'Guidance for non-poliovirus facilities to minimize risk of sample collections potentially infectious for polioviruses' (<u>http://polioeradication.org/wp-content/uploads/2018/04/polio-containment-guidance-for-non-poliovirus-facilities-20180410-en.pdf</u>) with or without a transfection reagent, except under the biorisk management conditions set-out in Annex 2 or Annex 3 of this standard.

Objectives

The objectives of this document are:

- to briefly described the Structure of the GPLN based on laboratory capacities
- to guide GPLN laboratories on handling and storage of biological materials containing type 2 polioviruses
- to establish standards and timeline for response to any polio events and/or outbreaks.

Audience

The proposed audience for this document is the Head and the personnel of laboratories members of the Global Polio Laboratory Network.

Reference documents

Additional information that may be useful to users of this document includes:

- Polio Laboratory Manual.¹ •
- The Global action Plan to minimize poliovirus facility-associated risk after type specific eradication of wild polioviruses and sequential cessation of OPV use (GAP-III)²
- Laboratory Biosafety Manual ^{3.} •

List of acronyms

AFP	Acute Flaccid Paralysis				
ES	Environmental Surveillance				
FTA®	Fast Technology for Analysis				
GPEI	Global Polio Eradication Initiative				
GPLN	Global Polio Laboratory Network				
ITD	Intratypic Differentiation				
LC	Laboratory Coordinator				
LQC	Laboratory Quality Control				
OPV	Oral Polio Vaccine				
NEV	Non Enterovirus				
NPEV	Non Polio Enterovirus				
NSL	Non Sabin like				
PEF	Poliovirus Essential Facility (for storage and handling of PV2)				
PV	Poliovirus				
RNA	Ribonucleic Acid				
rRTPCR	real time Reverse Transcriptase Polymerase Chain Reaction				
NIBSC	National Institute for Biological Standards and Controls. UK.				
SL	Sabin like				
SSECI	Stools, Sewage, Extracts, Concentrates and Isolates				
SOP	Standard Operating Procedure				
VDPV	Ambiguous vaccine-derived poliovirus				
VI	Viral Isolation				
VII	Viral Isolation and Identification				
VIIS	Viral isolation Identification and Sequencing				
WHO	World Health Organization				

¹ Polio Laboratory Manual and supplement at:

http://www.polioeradication.org/ResourceLibrary/GPLNpublications.aspx ² GAP-III. So far English and French are available at:

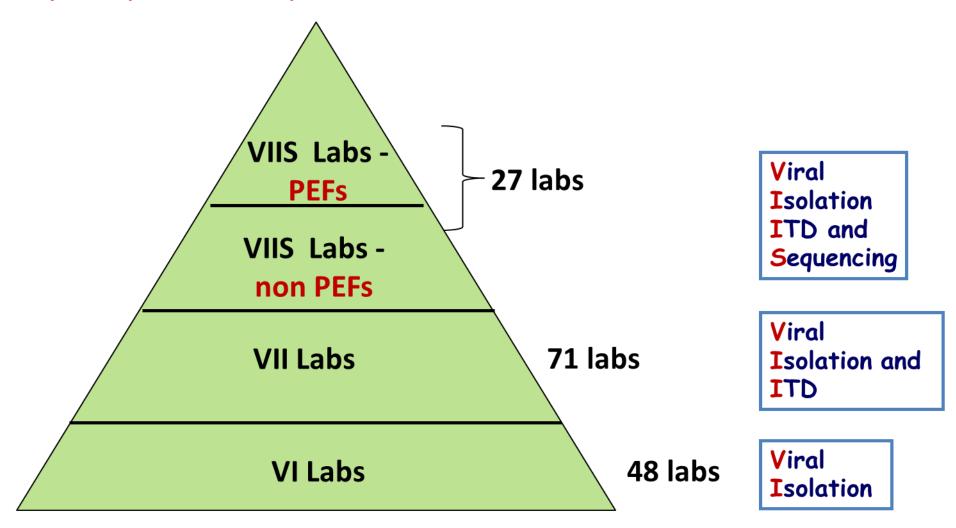
http://www.polioeradication.org/Posteradication/Containment.aspx . ³ Laboratory Biosafety Manual.

http://www.who.int/csr/delibepidemics/WHO CDS CSR LYO 2004 11/en/

Summary

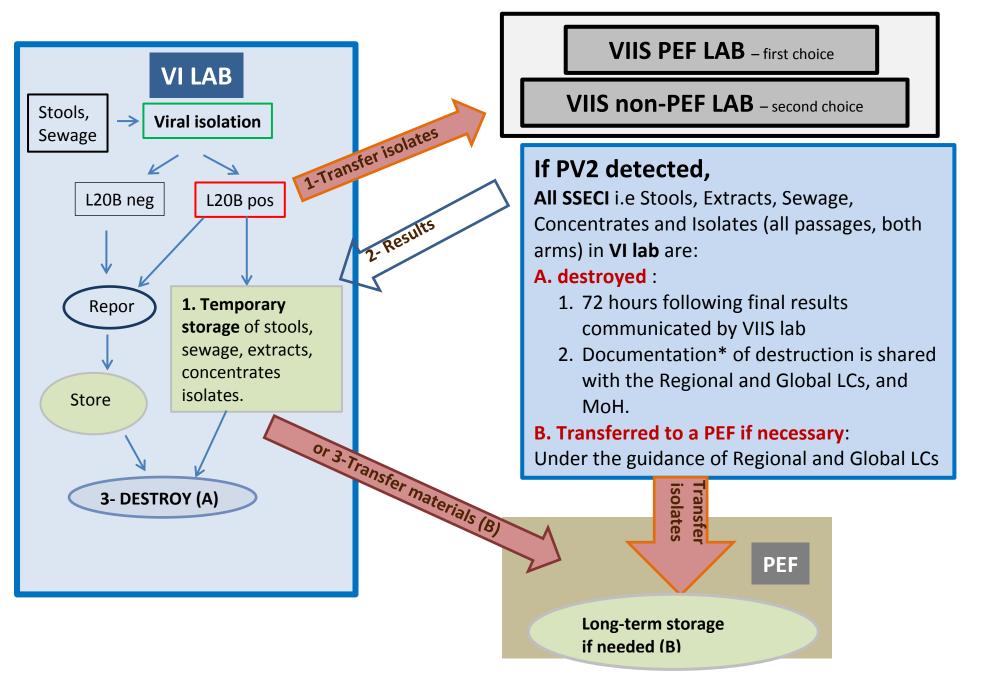
Viral isolation procedure Follow standard protocol. Timeline: 14 days	ITD procedure Follow standard protocol (on L+R+ or R+L+R+) Timeline: 7 days			
Suspected polioviruses (L+R+ or R+L+R+) Refer for ITD	PV2 negative: Proceed following standard algorithm.			
	 PV2 positive: 1. Report to Ministry of Health (MoH) and WHO in 24 hours 2. All original stool samples, stool extracts and cell-culture harvests to be packed, sealed and kept under lock and key at -20°C 3. Sent sample for sequencing and track. 4. When sequencing results are received (=day 0), immediate notification is sent to the Ministry of Health and WHO (acknowledgement of receipt needed). 5. Obtain green-light (at day 3 at the latest) and destroy the sealed package, document and share with MoH and WHO. 			

Capacity & Facility-based structure of the GPLN



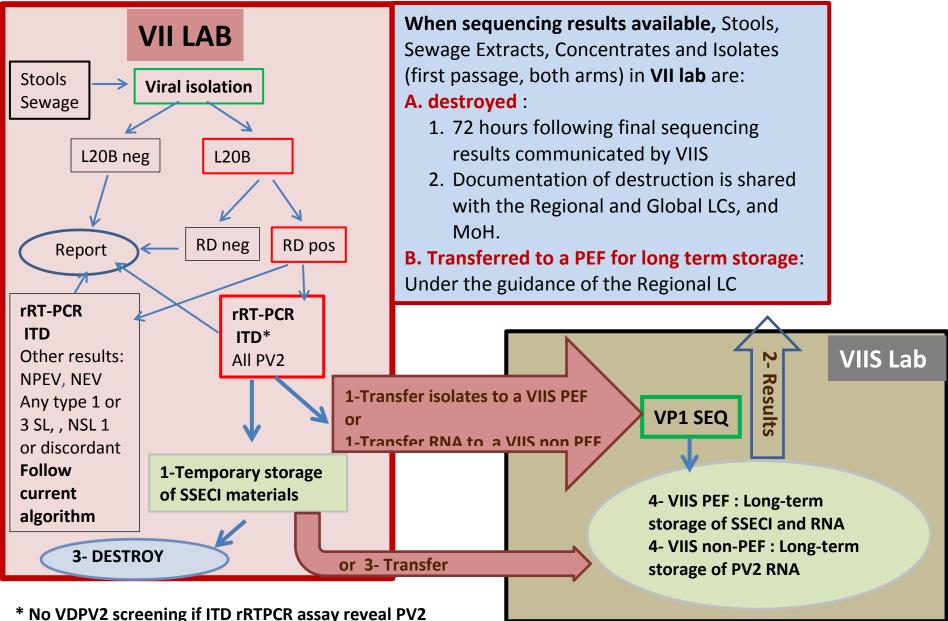
VIRAL ISOLATION (VI) LAB

SCHEME FOR BIOLOGICAL MATERIALS REFERRAL AND HANDLING



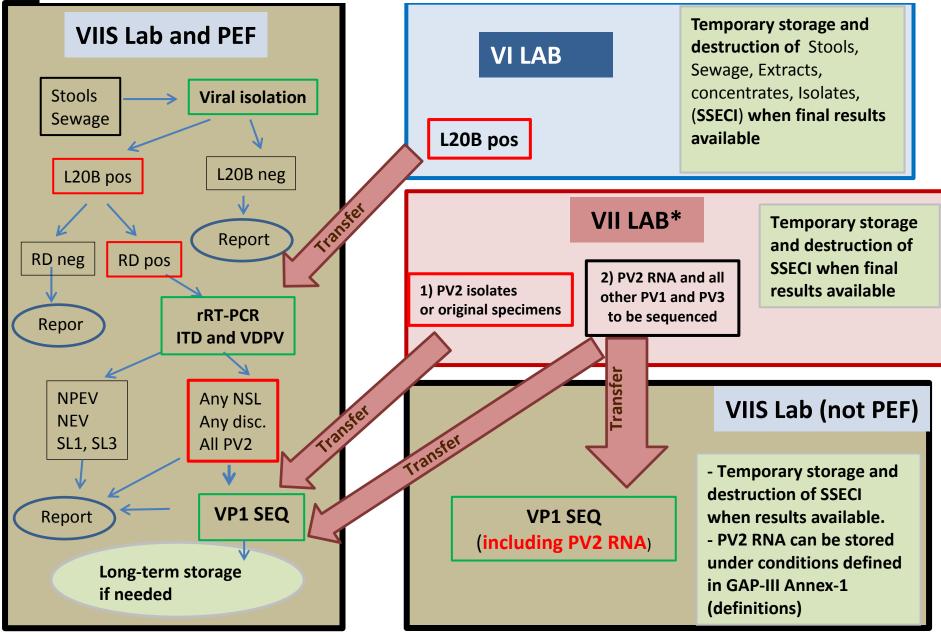
VIRUS ISOLATION AND IDENTIFICATION (VII) LAB

SCHEME FOR BIOLOGICAL MATERIALS REFERRAL AND HANDLING



VIRUS ISOLATION, IDENTIFICATION AND SEQUENCING (VIIS) LAB

SCHEME FOR BIOLOGICAL MATERIALS REFERRAL AND HANDLING



* No VDPV2 screening if ITD rRTPCR assay reveal PV2

Summary: handling and storing materials containing PV2

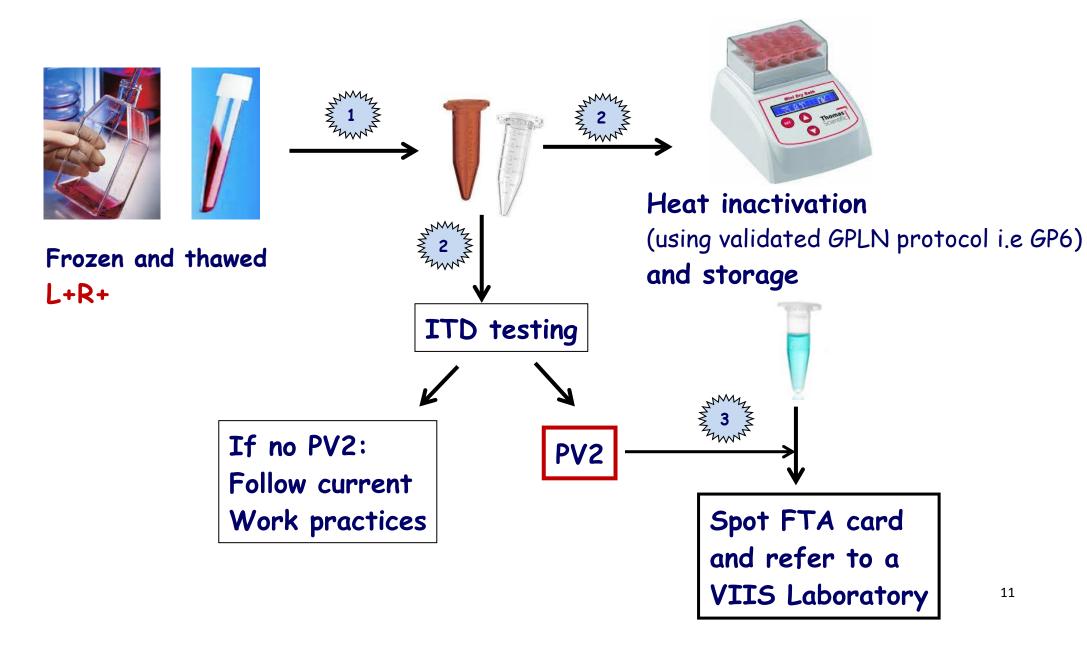
	Poliovirus Isolation (VI) Laboratory	Poliovirus Isolation and Identification (VII) Laboratory	Poliovirus Isolation, Identification and Sequencing (VIIS) Laboratory - Non PEF	Poliovirus Isolation, Identification and Sequencing (VIIS) Laboratory - PEF
Stool samples	Extraction + cell c.	Extraction + cell c.	Extraction + cell culture	Extraction + cell culture
Stool samples extracts				
Raw sewage samples	Concentration	Concentration	Concentration	Concentration
Concentrates of sewage samples	Cell-culture	Cell-culture	Cell-culture	Cell-culture
Suspected Poliovirus isolates (L+R+)		ITD rRT-PCR	ITD rRT-PCR	ITD rRT-PCR
Poliovirus isolates ITD result on L+R+ = PV2			PV inactivation and RNA extraction	RNA extraction
PV2 RNA from Polio isolates (L+R+).			sequencing	sequencing
NIBSC and LQC Sabin 2 Strains	Destroy (and document) by end of July 2016			CST, Serology

process Short-term storage until sequencing results available Long-term storage Destroy or transfer to an PV Essential Facility for storage

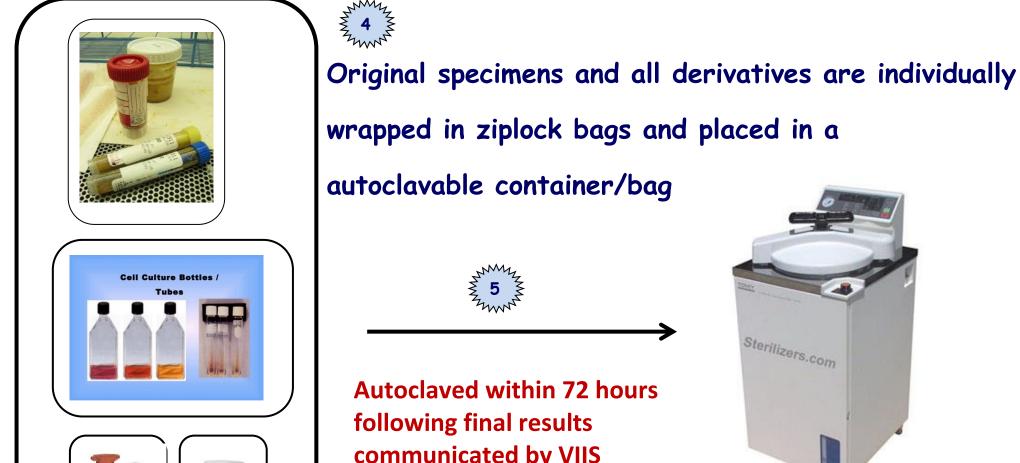
when sequencing results available (PV2)

Practical handling of biological materials containing PV2

Practical handling scheme (1) Aliquoting and testing



Practical handling scheme (2) Temporary storage and destruction



following final results communicated by VIIS laboratory (unless otherwise advised by WHO)

